The micrographs indicate that fibrin is produced through a predominantly lateral association of fibrinogen filaments. The characteristic striation does not appear under all circumstances, for example, in fibrils with widths below about 200 Å., or in tapered ends with widths less than this. Unstriated fibrils show a randomly particulate structure, while in the striated portions the particles are more concentrated in the stained bands. The micrographs of fibrinogen do not show any degree of regularity either in length or internal structure comparable to the regularity in fibrin. It appears that the periodicity in fibrin is not a manifestation of rigid dimensional units in fibrinogen, but is, rather, a characteristic developed The anomalsubsequent to initial aggregation ous variations in protein concentration indicate that some of the constituents have experienced local axial shifts to preferred positions.

Department of Biology

MASSACHUSETTS INSTITUTE OF TECHOLOGY

CAMBRIDGE, MASSACHUSETTS C. E. HALL RECEIVED FEBRUARY 12, 1949

FERMENTATION OF GLUCOSE-1-C¹⁴

Sir:

We have synthesized d-glucose-1-C¹⁴ from d(-)-arabinose by the Fischer-Kiliani method.¹ The mixed glucono- and mannonolactones were reduced catalytically,² carrier glucose added, and the radioglucose isolated and recrystallized to constant specific activity (about 6000 counts per gram minute). The glucose (in 0.2-g. samples) was then fermented anaerobically by Fleischmann's baker's yeast in phosphate buffers in the absence of a source of combined nitrogen; the fermentation yields (based on the carbon dioxide obtained) were 75-90%. The Embden-Meyerhof fermentation mechanism³ predicts that all the radiocarbon will appear in the methyl group of the alcohol so obtained; this prediction has been substantially confirmed.

The carbon dioxide obtained directly in the fermentation was counted as $BaCO_3$. The alcohol was degraded to acetone by the method outlined below; the latter was counted as its 2,4-dinitrophenylhydrazone.

$$\begin{array}{ccc} C^{14}H_3CH_2OH & \underbrace{K_3Cr_2O_7}_{H_2SO_4} & \underbrace{C^{14}H_3CO_2H}_{CH_3CO_2H} & \underbrace{Ba(OH)_2}_{H_2SO_4} \\ & (C^{14}H_3CO_2)Ba(O_2CCH_3) & \underbrace{530^{\circ}}_{Vacuum} \\ & C^{14}H_3COCH_3 + BaCCC \end{array}$$

In a set of control experiments on the pyrolysis using methyl-labelled acetic acid,⁴ we found 0.1-0.3% of the activity in the barium carbonate produced. Further in these control experiments, the

(1) Kiliani Ber., 19, 3033 (1886).

(3) Meyerhof, Biochem. Symposia, ∇, 141 (1941).

(4) The methyl labelled acetic acid was kindly supplied to us by Professor Konrad Bloch

specific activity of the acetone 2,4-dinitrophenylhydrazone was only 77% of that anticipated from the specific activity of the barium acetate, even after making the usual corrections⁵ for self-absorption, etc. The specific activities of all samples of acetone 2,4-dinitrophenylhydrazone were therefore corrected by the factor 1/0.77. The final results, together with the counting errors (95% confidence level), are given below:

	Other	Per cent. of radioactivity (based on glucose fermented) found in		
¢Η	conditions	CO_2	CH₃	CH₂OH
6.2	Live yeast	6.5 ± 2.4	92 ± 4.0	4.8 ± 2.4
5.7	Live yeast	1.0 ± 1.8	76 ± 4.0	7.6 ± 2.8
5.7	Dried yeast			
	powder	3.7 ± 1.8	92 ± 7.8	0.2 ± 0.4

We are currently investigating the causes of the slight radioactivity in the carbon dioxide obtained directly in the fermentation and in the barium carbonate from the pyrolysis.

(5) Yankwich and Weigl, Science, 107, 651 (1948): Libby, Ind. Eng. Chem., Anal. Ed., 19, 2 (1947).

GEORGE HERBERT JONES LABORATORY

THE UNIVERSITY OF CHICAGO DANIEL KOSHLAND, JR. CHICAGO, ILLINOIS F. H. WESTHEIMER RECEIVED FEBRUARY 3, 1949

A NEW SYNTHESIS OF 2-PHENAZINOL, THROUGH THE DI-N-OXIDE

Sir:

The chlorine atom of 2-chlorophenazine can be labilized toward aqueous-alcoholic sodium or potassium hydroxide by converting the base to 2chlorophenazine-5,10-dioxide (red-orange needles, m. p. 190–191° (dec.)¹ Anal.² Calcd. for $C_{12}H_{7}$ -ClN₂O₂: C, 58.4; H, 2.87. Found: C, 58.5; H, 3.06. This conversion is readily effected, using the method employed by Clemo and McIlwain³ for other phenazines. Refluxing the chlorophenazine dioxide for about twelve hours with aqueousalcoholic potassium or sodium hydroxide gives a deep purple solution of the alkali salt, from which hydrochloric acid precipitates the free 2-phenazinol-5,10-dioxide (orange-red, begins to darken at 236°). Anal. Calcd. for $C_{12}H_8N_2O_3$: C, 63.2; H, 3.54. Found: C, 63.1; H, 3.87. Reduction of this by sodium hyposulfite (sodium "hydrosulfite") in alkaline solution at room temperature gives a red solution of the sodium salt of 2-phenazinol, from which the free phenol⁴ is precipitated by acid. (Clemo and McIlwain³ found sodium hyposulfite effective in reducing 1-phenazinol-5,10dioxide.) The 2-phenazinol may be purified by vacuum sublimation and chromatographic adsorp-

(1) All melting points are corrected.

(2) Analyses by Mr. W. C. Alford, Mrs. M. M. Ledyard and Mrs. E. G. Peake.

(3) Clemo and Mcllwain, J. Chem. Soc., 483 (1938).

⁽²⁾ Glattfeld and Schimpf, THIS JOURNAL, 57, 2204 (1935).

⁽⁴⁾ Kehrmann and Cherpillod, *Helv. Chim. Acta*, **7**, 975 (1924). As these authors indicate, in saying that the 2-phenazinol melts at "about" $253-254^{\circ}$ with decomposition, this compound does not appear to have a sharp melting point.